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## **Bispecifics Resistance and Combinations in B-Cell non-Hodgkin Lymphoma**

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- Overview of CD20xCD3 bispecific antibodies (BsAb) and mechanisms of resistance (MoR)
- Individual MoR and strategies to overcome:
  - Antigen loss
  - T-cell dependent mechanisms
  - Other

### **Overview of CD20xCD3 bispecific antibodies**

Product name	Schematic depiction	Format	Technology	CD20:CD 3 ratio	Approved indication(s)*	ORR (CR) , %	PFS (months)
Mosunetuzumab	CD20 CD3	lgG1	Knobs-into- holes (different Fabs)	1:1	R/R FL	80 (60)	17.9
Glofitamab	CD20 CD20 CD20	lgG1	Head-to-tail fusion	2:1	R/R DLBCL	52 (39)	4.9
Epcoritamab	CD20 CD3	lgG1	Controlled Fab- arm exchange	1:1	R/R DLBCL	63 (39)	4.4
					R/R FL	82 (62)	15.4
Odronextamab	CD20 CD3	lgG4	Heavy chains with different affinity	1:1	R/R DLBCL	52 (31)	not rep
					R/R FL	80 (73)	20.7

\* Variably approved by the U.S. FDA, EMA and other regulatory agencies

Falchi L et al. *Blood.* 2023;141(5):467-480; Budde LE et al. Lancet Oncol. 2022;23(8):1055-1065; Dickinson MJ et al. *N Engl J Med.* 2022;387(24):2220-2231; Thieblemont C et al. J Clin Oncol 2023;41:2238-2247(2023); Linton K et al. Lancet Haematol 2024;11(8):e593-e605; Kim TE et al. Ann Oncol 2024 (ahead of print); Ayyappan S et al. *Blood* 2023; 142 (s1): 436.

## **Basic mechanism of action of bispecific antibodies**







### **Overcoming resistance to CD20xCD3 bispecific antibodies**



## Mechanism of resistance #1 Antigen (CD20) loss

# Mosunetuzumab in R/R FL: CD20 loss is frequent and associated with progressive disease





# Post-mosunetuzumab CD20 loss is only partly explained by *MS4A1* mutations



*MS4A1* mutations were pre-existent in 10/154 patients with CD20 expressed by IHC in all cases



Detection of CD20 by flow cytometry in SU-DHL-16-engineered cell lines.

- Intracellular expression detected after permeabilization using anti-CD20 antibody targeting the C terminus (H-1, BD-561174),
- Extracellular expression detected using anti-CD20 antibody targeting ECL2 (2H7, BD-555623)

## CD20 loss in patients with B-NHL treated with epcoritamab may be explained by post-transcriptional modifications of *MS4A1* and mechanisms differ in CR vs. PD







Pre-T

On-Tx

# Working around CD20 loss: JNJ-80948543 (CD79bxCD20xCD3) trispecific Ab



#### Broader antigen targeting (T-cells from healthy donors)



### Effect on Tumor Cells Expressing CD79b and/or CD20



T-cell Redirection Assays; E:T ratio 5:1

Note: No clinical data released to date.

## Mechanism of resistance #2 Qualitative and quantitive T-cell changes

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# On-treatment changes in T-cell subsets govern responses in patients with B-NHL treated with epcoritamab





Large (21-100) • Medium (6-20) • Small (2-5) • Single



## CD8+ cytotoxic T-cells specifically mediate epcoritamabinduced cell killing





## Restoring T-cell dysfunction: glofitamab + RO7443904 (CD19xCD28 co-stimulatory Ab)



Humanized NSG mice (7-8mice/group) implanted with WSU-DLCL2 Fluc lymphoma cells i.v. and treated once weekly i.v. with glofitamab (0.15mg/kg) or CD19-CD28 (1mg/kg) in monotherapy or with the combination starting on day 3. Tumor growth was recorded with BLI measurement.



Humanized NSG mice were inoculated s.c. with WSU-DLCL2 lymphoma cells and received obinutuzumab followed by glofitamab (0.15mg/kg) alone or with CD19-CD28 (1mg/kg). Tumors were harvested 8 days later



IF in tumors for CD8+CD28+ human T cell frequencies at 6 days after treatment with glofitamab alone or with CD19-CD28(1mg/kg)

# Englumafusp alfa (CD19x4-1BBL) + glofitamab in R/R aggressive B-NHL (N=83)





- Population: Median age 63; prior lines = 3 (1-8); refractory to CART N=35
- **Safety:** CRS 55% (G3 1%); infections 58% (G5 5%); neutropenia 25% (G3 20%)
- Efficacy: ORR 67%; CR 57%; CR post CAR-T 48%

### Lenalidomide as rational combination partner for epcoritamab



### **R<sup>2</sup>-epcoritamab in High-Risk R/R FL: Responses and Outcomes**





# Insights into SC mosunetuzumab in 1L FL: Hints to mechanisms of resistance



# BTK inhibitors enhance CD8+ tumor response and may synergize with bispecific antibodies

Serial peripheral blood samples from patients treated in study E1912



### Ibrutinib + rituximab



Ibrutinib + glofitamab

- %CD4+ or CD8+ T-cell:CLL conjugates at baseline, 6M, and 12M (n = 15).
- Representative confocal images of CD8+ (white) and CD4+ (green) T-cell conjugates with CLL B cells (blue)
- T-cell-mediated CLL cell death using purified T cells from B/L, 6M, 12M, or 18M ibrutinib-rituximab time points against target B/L CLL B cells after ex vivo treatment with glofitamab (0.01 µg/mL) or nonbinding antibody control

## Mosunetuzumab + Zanubrutinib in 1L FL: Study design



PET/CT, positron emission tomography/ computerized tomography; CR, complete response; PR, partial response; SD, stable disease; PD, progressive disease; <sup>b</sup> CT must be of diagnostic quality; <sup>a</sup> Patients who experience PD will be taken off study. A biopsy to confirm PD is recommended.

## Mechanism of resistance #3 Non-T-cell microenvironmental changes: No data!

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## IPH6501, first-in-class tetraspecific NK-engager targeting CD20 for the treatment of B-NHL



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Gauthier L et al. Cell. 2019;177(2):1701-1713. Colomar-Carando N et al. Cancer Immunol Res. 2022;10(3):291-302. Gauthier L et al. Nature Biotech. 2023;41:1296-1306. Demaria O et al. Cell Rep Med. 2022;3(10):100783; Demaria O et al. Science Immunol, in press.

### **Overcoming resistance to CD20xCD3 bispecific antibodies**



## Take-home messages and open questions

- BsAb are a critically important addition in R/R B-NHL
  - Consistent efficacy with predictable safety
  - The majority of patients experience progression / relapse

### Resistance to BsAb is multifaceted

- CD20 loss is highly prevalent and associated with BsAb failure
- Many unknowns in T-cell dependent resistance (timing, topography, reversibility)
- No information on the role of other immune cells

### Strategies to overcome resistance are emerging

- High response rates with emerging combinations
- Is CD20 reversible in some cases?
- Should T-cell subsets be differentially leveraged for optimal efficacy (rather than pan-T-cell strategies)?

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